

Inhibiting IDH mutations in low-grade glioma alters cellular function and the immune environment

Min Lu¹, Ingo K Mellinghoff², Aaron Diaz³, Jennie W Taylor³, Sung Choe¹, Ania Tassinari¹, Dongwei Zhu¹, Kha Le¹, Feng Tai¹, Islam Hassan¹, Shuchi S Pandya¹, Lori Steelman¹, Bin Wu¹

¹Agios Pharmaceuticals, Inc., Cambridge, MA, USA; ²Memorial Sloan Kettering Cancer Center, New York, NY, USA; ³University of California San Francisco, San Francisco, CA, USA

BACKGROUND

Isocitrate dehydrogenase (IDH) mutations in cancer

- Somatic mutations in the metabolic enzyme IDH occur in many cancers, with mutations in *IDH1* and *IDH2* occurring in approximately 80% and 4% of lower-grade gliomas (LGG; WHO grade 2/3), respectively.^{1,2}
- Mutant IDH (mIDH) proteins have a gain-of-function enzyme activity, catalyzing the reduction of alpha-ketoglutarate to the oncometabolite D-2-hydroxyglutarate (2-HG)^{3,4}
 - 2-HG inhibits alpha-ketoglutarate-dependent enzymes, resulting in epigenetic dysregulation, impaired cellular differentiation, and oncogenesis^{5,6}
- In preclinical models of leukemia, glioma, and sarcoma, inhibitors of mIDH enzymes blocked 2-HG production and showed antitumor activity^{7,8}
- IDH* mutations are associated with immune evasion in gliomas
 - mIDH* gliomas exhibit fewer tumor-infiltrating lymphocytes and reduced protein expression of programmed death ligand 1 (PD-L1) than wild-type (WT) counterparts⁹⁻¹⁵
 - Mechanistically, 2-HG is immunosuppressive and plays a key role in modulating the tumor immune microenvironment in *mIDH* gliomas¹⁶⁻¹⁸
 - mIDH inhibition in combination with vaccine therapy or PD-L1 blockade increased tumor-infiltrating lymphocytes and improved survival in preclinical glioma models^{19,20}

Inhibitors of mIDH

- Ivosidenib (IVO; AG-120)** is a first-in-class, oral, small-molecule inhibitor of mIDH1
 - Approved by the US FDA for *mIDH1* relapsed/refractory acute myeloid leukemia and in a subset of patients with *mIDH1* newly diagnosed acute myeloid leukemia
 - In an ongoing phase 1 study including 66 patients with glioma, IVO was associated with a favorable safety profile and clinical activity at 500 mg once daily (QD)
 - Objective response rate of 5.7% (including 1 minor response) and 83% stable disease rate, with a median progression-free survival of 13 months in non-enhancing glioma (n = 35)²¹
- Vorasidenib (VOR; AG-881)** is an oral, potent, reversible, brain-penetrant, pan-inhibitor of mIDH1 and mIDH2
 - In an ongoing phase 1 study including 52 patients with glioma, VOR was associated with a favorable safety profile and clinical activity at doses < 100 mg QD
 - Objective response rate of 18.2% (including 1 partial response and 3 minor responses) and 73% stable disease rate, with a median treatment duration of 25.8 months and a median progression-free survival of 31.4 months in non-enhancing glioma (n = 22)²²
 - In an ongoing perioperative study (ClinicalTrials.gov NCT03343197), IVO and VOR demonstrated brain penetration and > 90% suppression of 2-HG in resected *mIDH1* gliomas after preoperative treatment for approximately 4 weeks²³

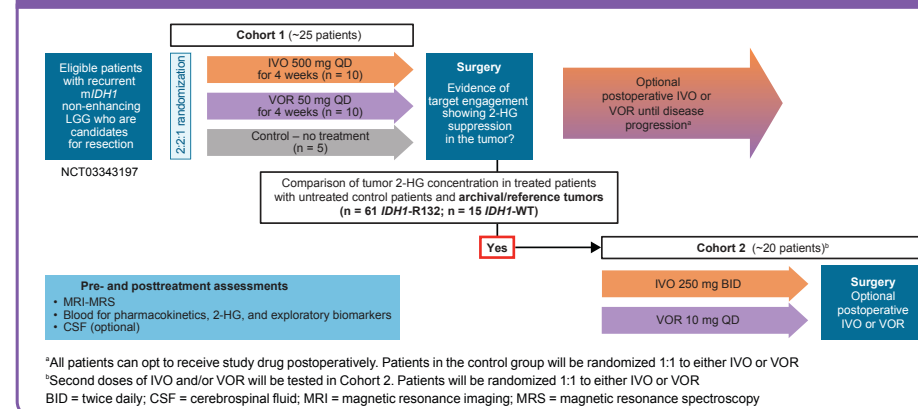
OBJECTIVES

- To understand the molecular and cellular mechanisms underlying mIDH inhibition by IVO or VOR in LGG using resected tumor tissues from patients with glioma enrolled in a phase 1 perioperative study (ClinicalTrials.gov NCT03343197)

METHODS

- The study schema for the phase 1 glioma perioperative study is shown in **Figure 1**

Figure 1. Study design



Sample collection

- Fresh frozen tissue samples included a set of banked reference *IDH* WT and *mIDH* LGG samples, as well as *mIDH* samples collected from enrolled patients who underwent surgery
- Formalin-fixed paraffin-embedded samples included paired archival (pretreatment) and surgical (posttreatment) samples from enrolled patients
- The actual number of patients with available data is noted in each figure

METHODS (CONTINUED)

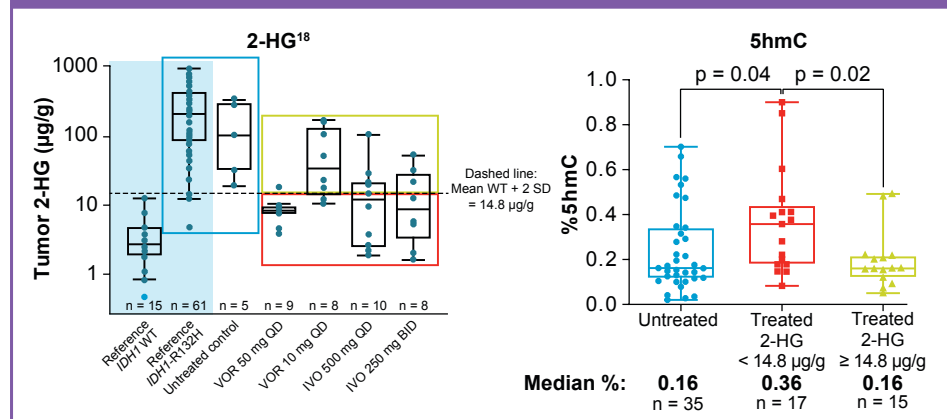
Biomarker assessment

- Next-generation sequencing: DNA and RNA samples were extracted from fresh frozen tumor tissues, and DNA sequencing and RNA sequencing were conducted at Personalis (Menlo Park, CA)
- 5-hydroxymethylcytosine (5hmC) analysis: DNA samples were digested with DNA degradase (Zymo Research, Irvine, CA) to generate single nucleosides; 5hmC, 5-methylcytosine, and cytosine were later quantified using liquid chromatography-tandem mass spectrometry
- RNA sequencing analysis: mRNA expression levels in transcripts per million (TPM; raw counts corrected for gene length and sequencing depth) were compared between untreated samples and samples treated with IDH inhibitors, and the significance of the difference was assessed using the Wald test
- T-cell receptor (TCR) sequencing was performed at Life Technologies Clinical Services Lab (Sacramento, CA) using OncoPrint TCR Beta-SR Assay with RNA samples extracted from fresh frozen tissue
- Immunohistochemistry (IHC) for Ki-67, CD3, and CD8 was performed by Mosaic Laboratories (Lake Forest, CA), and quantification derived from an annotation including all tumor and intervening stroma within the tumor nest
- Multiplex IHC for CD68, CD163, and HLA-DR was carried out at Akoya Biosciences (Marlborough, MA), and image analysis performed using Phenochart software

RESULTS

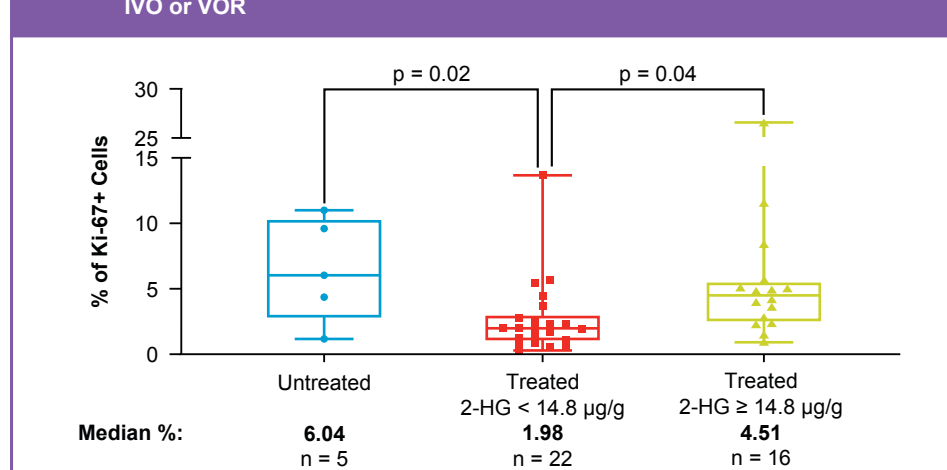
- mIDH inhibition increased 5hmC, a marker of DNA demethylation, suggesting reversal of DNA hypermethylation (**Figure 2**)
 - Optimal 2-HG suppression (defined as posttreatment 2-HG levels < 14.8 µg/g, which is the upper range of *IDH* WT tumor 2-HG levels) in IVO- or VOR-treated samples (n = 17) led to a ~2-fold increase in 5hmC compared with untreated controls and banked reference samples (n = 35; p = 0.04)

Figure 2. 2-HG and 5hmC levels following mIDH inhibition with IVO or VOR



- IHC analysis of the proliferation marker Ki-67 showed a ~3-fold decrease in the percentage of Ki-67-positive cells in IVO- or VOR-treated samples with optimal 2-HG suppression (n = 22) compared with untreated controls (n = 5; p = 0.02) (**Figure 3**)

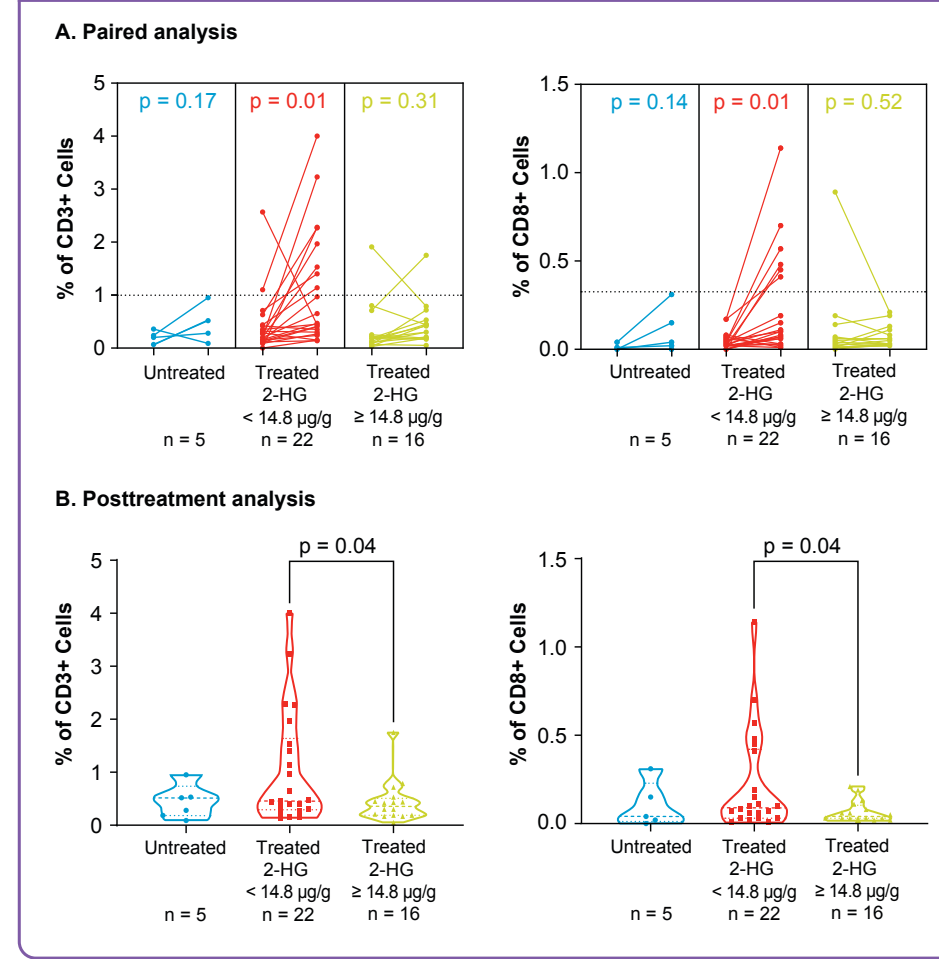
Figure 3. Tumor cell proliferation assessment by IHC following mIDH inhibition with IVO or VOR



RESULTS (CONTINUED)

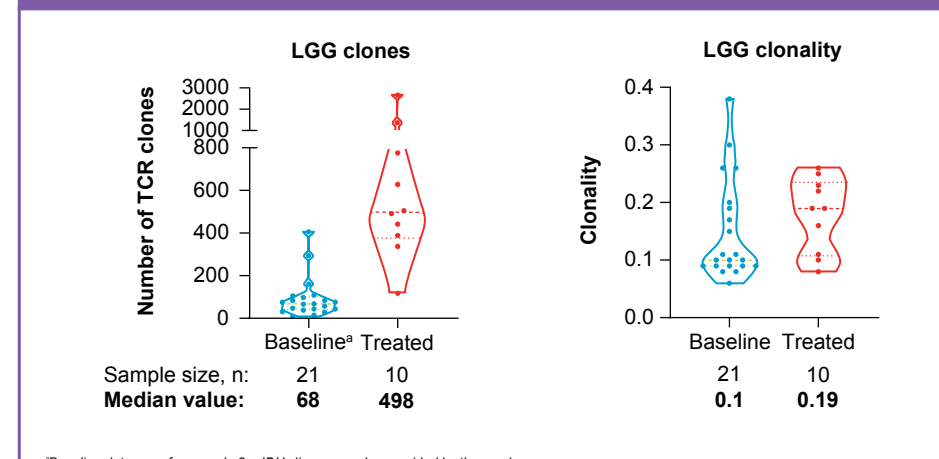
- IHC analysis from paired pretreatment (archival) and posttreatment samples showed an increase of CD3+ T-cell infiltration (p = 0.01) and CD8+ T-cell infiltration (p = 0.01) in IVO- or VOR-treated samples with optimal 2-HG suppression (n = 22) (**Figure 4A**)
- IHC analysis from posttreatment samples also showed an increase of CD3+ and CD8+ T-cell infiltration in IVO- or VOR-treated samples with optimal 2-HG suppression compared with untreated controls (**Figure 4B**)

Figure 4. T-cell infiltration assessment by CD3 IHC and CD8 IHC following mIDH inhibition with IVO or VOR



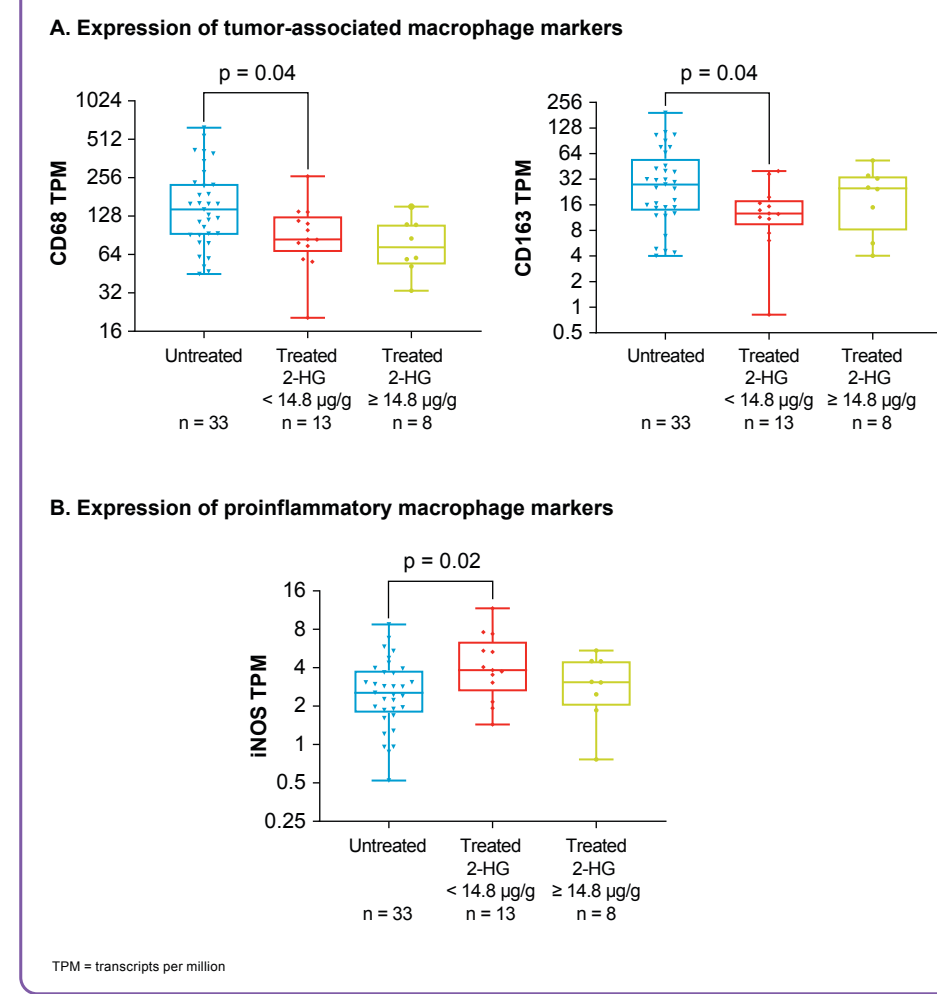
- mIDH inhibition increased the number of unique TCR clones as well as clonality assessed by TCRB sequencing (**Figure 5**)

Figure 5. TCR clones and clonality assessment by TCRB sequencing following mIDH inhibition with IVO or VOR



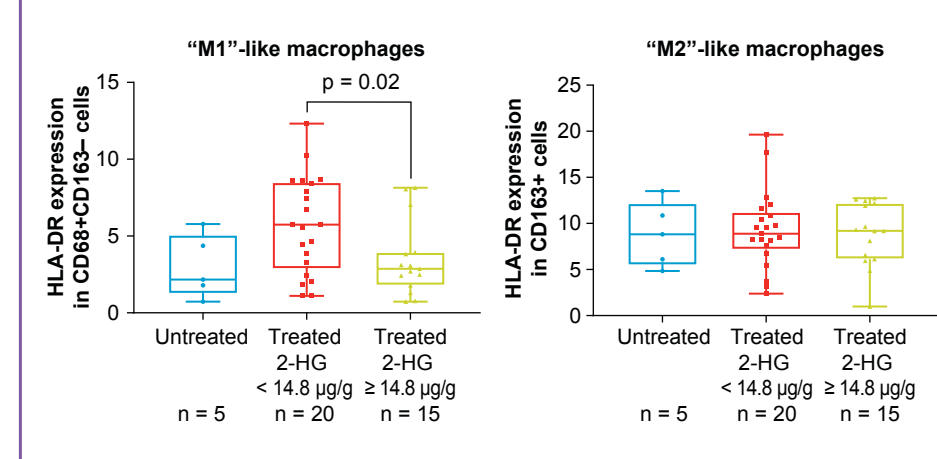
- IDH inhibition decreased the expression of tumor-associated macrophage marker genes and increased the expression of proinflammatory macrophage marker genes in *mIDH* oligodendroglioma samples with optimal 2-HG suppression (**Figure 6**)

Figure 6. Expression of macrophage markers following mIDH inhibition with IVO or VOR



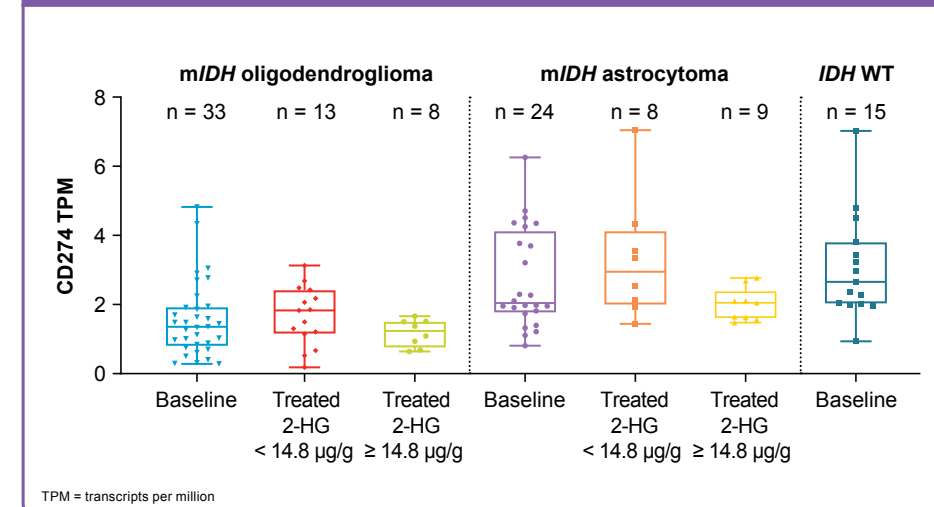
- mIDH inhibition increased the expression of HLA-DR in "M1"-like macrophages (p = 0.02) but not in "M2"-like macrophages in samples with optimal 2-HG suppression assessed by IHC (**Figure 7**)

Figure 7. Expression of HLA-DR following mIDH inhibition with IVO or VOR



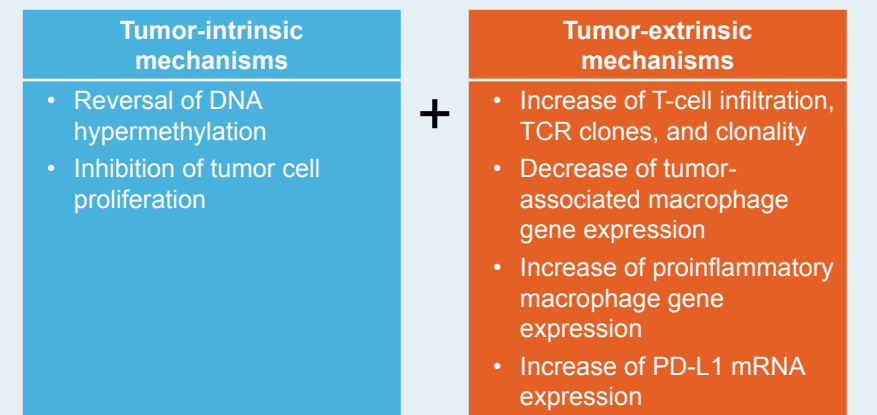
- mIDH inhibition upregulated PD-L1 mRNA expression in samples with optimal 2-HG suppression (**Figure 8**)

Figure 8. Expression of PD-L1 mRNA following mIDH inhibition with IVO or VOR



CONCLUSIONS

- Overall, these findings suggest that both tumor-intrinsic and tumor-extrinsic mechanisms underlie mIDH inhibition by IVO or VOR treatment



- These data support further development of mIDH inhibitors in *mIDH* LGG either as a monotherapy or in combination with immunotherapy

Questions? Please contact Min Lu, Min.Lu@agiopharm.com.

Acknowledgments
We would like to thank the patients, families, coinvestigators, and all study personnel who participated in this study.

Disclosures
This study was funded by Agios Pharmaceuticals, Inc. ML, SC, AT, DZ, KL, FT, IH, SSP, and BW: Agios - employment and stockholder. IKM: Agios, Amgen, Bristol-Myers Squibb, General Electric, and Lilly - research funding; Agios, AstraZeneca, Black Diamond Therapeutics, DC Europa, Debiopharm, Puma Biotechnology, Voyager Therapeutics - research consultant/advisor; Roche - honoraria; AD: no conflicts of interest to disclose. JW: AbbVie, Agios, and Bristol-Myers Squibb - research funding; LS: Agios - employment and stockholder; Infinity Pharmaceuticals - stockholder. Editorial assistance was provided by Vanessa Duca, PhD, Excel Medical Affairs, Fairfield, CT, USA, and supported by Agios.

References

- Yan H et al. *N Engl J Med* 2009;360:765-73.
- The Cancer Genome Atlas Research Network. *N Engl J Med* 2015;372:2481-98.
- Dang L et al. *Nature* 2009;462:739-44.
- Ward PS et al. *Cancer Cell* 2010;17:225-34.
- Lu C et al. *Nature* 2012;483:474-8.
- Saha SK et al. *Nature* 2014;513:110-4.
- Xu W et al. *Cancer Cell* 2011;19:17-30.
- Wang F et al. *Science* 2013;340:622-6.
- Rohle D et al. *Science* 2013;340:626-30.
- Kohanchash G et al. *J Clin Invest* 2017;127:1425-37.
- Amankulor NM et al. *Genes Dev* 2017;31:774-96.
- Bunsee L et al. *Nat Med* 2018;24:1192-203.
- Friedrich M et al. 2018 SNO Annual Meeting; Abstr IMMUL-52.
- Zhang L et al. *Clin Cancer Res* 2018;24:5381-91.
- Berghoff AS et al. *Neuro Oncol* 2017;19:1460-8.
- Mellinghoff I et al. 2017 SNO Annual Meeting; Oral presentation ACTR-46.
- Mellinghoff I et al. 2020 ASCO Annual Meeting; Oral presentation 2504.
- Mellinghoff IK et al. 2019 SNO Annual Meeting; Oral presentation ACTR-66.